Project Stage	General Topic	Specific Metric(s)	Analysis Already Agreed To By USAF?
Pre-Baseline			
	Monitoring Well Installations		
		Continuous logging PID readings	Y
		LNAPL Dye Test; VOC and TPH if Dye Test is Positive	Y
		VOCs	Υ
		TPH (DRO, GRO)	Υ
Baseline Data		VOCs	

Timing of Analyses	Frequency of Analyses	Location of Analyses
Before baseline geochemistry, field data, and microbial analyses performed	(Once - is an installation)	(Location of Installations)
	Once	CZ
	Once	UWBZ
	Once	LSZ
during well installation		Following Table 5.1
during well installation		Following Table 5.1
during well installation		Following Table 5.1
		Following Table 5.1
		Following Table 5.1

Purpose
These are additional wells to provide accurate monitoring of EBR
These MWs are needed to ensure that there are sufficient data to evaluate the effectiveness of EBR.
The extraction wells can be used, but must be considered in separate groups and are not sufficient for this evaluation.
To determine if benzene is slower to degrade than other aromatics
(or faster, or average)

To provide one singular, synoptic round of data prior to inception of EBR

#### **Additional Comments**

MWs are needed in suitable locations to monitor the effectiveness of EBR. Otherwise, data evaluation will be much less meaningful. Accurate delineation of concentrations in downgradient portions of the site should also be emphasized relative to off-site migration potential, sulfate utilization, etc.

To the degree possible, wells should also be located so that aquifer heterogeneities (low-permeability zones) can be monitored and accurate spatial averages for parameter values can be computed.

New MWs must have time to equilibrate after installation and development before baseline field data, geochemistry, and microbial analyses are performed.

7 treatment "ovals" proposed, but only 3 ovals have monitoring wells that are in reasonable locations. Monitoring wells should be installed in locations between the injection and extraction wells to evaluate sulfate distribution and EBR progress (5/II/17 BCT slides, slide 25)

5 initial treatment "ovals" proposed; however, only one of the first 5 "ovals" where EBR is proposed for initial implementation has a monitoring well (ST012-UWBZ24). This well is not located in an optimal location for monitoring the effectiveness of treatment (i.e., it is not located on the path between the injection and extraction wells). Since these ovals are proposed for the initial injections, at least one monitoring well should be installed in each oval treatment area so that the injections and EBR progress can be monitored. There are 5 additional treatment "ovals," but there are no monitoring wells in these ovals; monitoring wells should be installed (5/11/17 BCT slides, slide 26)

15 treatment "ovals" proposed, but only 2 have monitoring wells in suitable locations. 3 additional "ovals" have monitoring wells located beyond the extraction well. Depending on how the extraction wells are pumped, sulfate may never reach these monitoring wells. Monitoring wells should be installed in locations that are suitable to monitor injections and EBR progress. The wells located beyond the extraction wells should also be monitored to evaluate sulfate distribution (5/11/17 BCT slides, slide 27)

Taken from Table 5.1, RD-RAWP Addendum 2 (March 2016)

Taken from Table 5.1, RD-RAWP Addendum 2 (March 2016)

Taken from Table 5.1, RD-RAWP Addendum 2 (March 2016)

Taken from Table 5.1, RD-RAWP Addendum 2 (March 2016)

Taken from Table 5.1, RD-RAWP Addendum 2 (March 2016)

These data, collectively, will help establish baseline criteria against which project progress and goals can be compared and monitored.

# **Hydrogeologic Data**

Groundwater gauge data (depth to	
water, depth to product, product	
thickness)	
Perform Slug Tests	

# **Mapping Contaminant Locations and Concentrations**

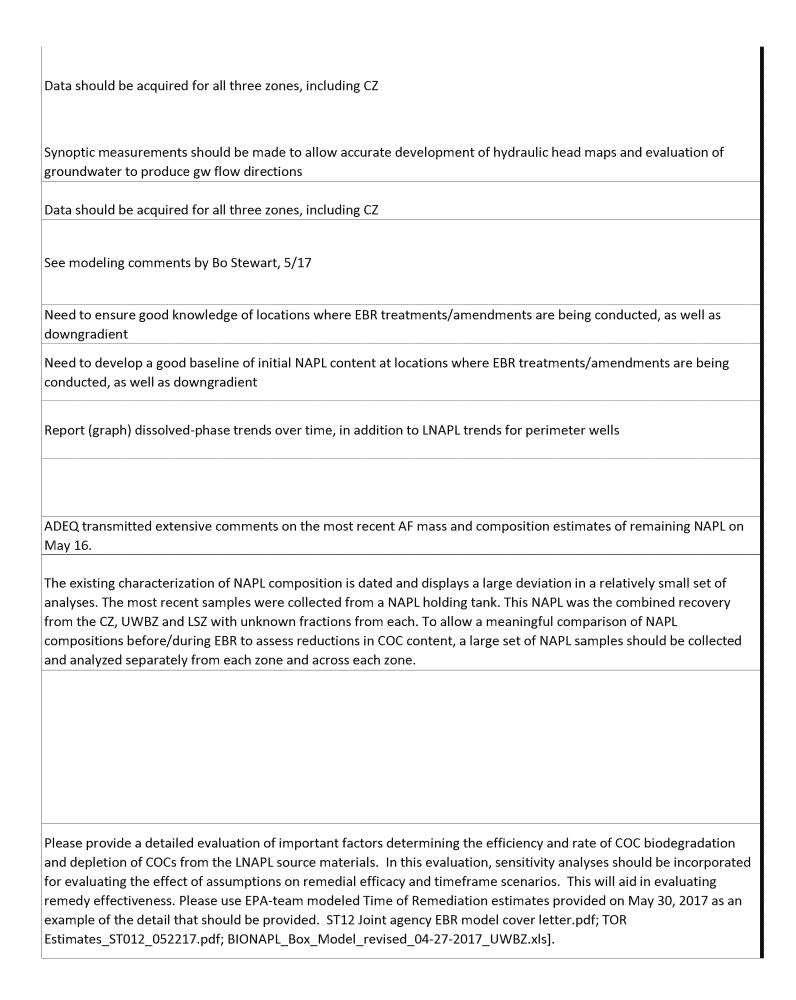
Continue to locate and map LNAPL presence and depth	Υ
Monitor benzene content and concentration in LNAPL, where LNAPL is found	Y
Continue to locate and map dissolved- phase benzene presence and concentration	Y
Continue to locate and map dissolved- phase VOC presence and concentration Calculate total LNAPL mass present at start of EBR	Υ
Determine the content of COCs in the LNAPL at the start of EBR	
Locate and map sulfate concentrations	Y

# Modeling

and soil
----------

After SEE but before EBR injections or amendments	Once as baseline	New and existing MWs, located in the area to be impacted by injections/amendments, and downgradient of this area
		All New Wells and Existing Wells that have not been tested
After SEE but before EBR injections or amendments	Once as baseline	New and existing MWs, located in the area to be impacted by injections/ amendments, and downgradient of this area
	monthly	Perimeter wells
		New and existing MWs with recoverable LNAPL
		Targeted treatment area and downgradient portions of the site
After SEE but before EBR injections or amendments	Once as baseline	

For use in modeling
Hydraulic Conductivity Measurement; for use in modeling
Refer notes in "modeling" section of this table.
Comparison of NAPL compositions before/during EBR to assess reductions in COC content
When compared to this baseline data, this information will help monitor for sulfate migration outside of the COC areas and facilitate comparison of EBR modeling results with field data



Provide details of EBR modeling to calculate time estimates for remediation	
Provide proof of concept supporting the sulfate reduction for EBR	
Provide details used to determine the optimal sulfate injection strategy.	

# **GW Geochemistry**

Tomporaturo	γ
Temperature	•
pH	Υ
ORP value	Υ
Dissolved Oxygen	Υ
Nitrate	Υ
Phosphorus	
Ferrous Iron	
Total Iron	
Sulfate	Υ
Hydrogen Sulfide	
Methane	
Alkalinity	
TPH (DRO, GRO)	Υ
VOCs	Υ
Arsenic	Υ

# **Indigenous Microbial Population**

Total size	
Major groups within population, and	
their proportion of total	

		T
After SEE but before EBR injections or amendments	Once as baseline	New and existing MWs, located in the area to be impacted by injections/ amendments, and downgradient of this area
After SEE but before EBR injections or	Once to establish	Samplers should be placed so as to monitor the core of sulfate injections, its periphery, and downgradient.
amendments	baseline	All three zones should be monitored.  The same wells should be monitored pre-EBR, during EBR, and post-EBR.

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There are the contiller considerable size on the continue of the class	1
These analyses will quantify the size, makeup, and health of the	
indigenous microbial community.	
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Please use EPA-generated models as an example of sufficient documentation, to allow for an independent evaluation of the results of the modeling. Please see comments provided in: Techlaw memo dated March 24, emailed on April 11, 2017, and Time of Remediation estimates provided on May 30, 2017 (ST12 Joint agency EBR model cover letter.pdf; TOR Estimates_ST012_052217.pdf; BIONAPL_Box_Model_revised_04-27-2017_UWBZ.xls])
In particular, very little field data exists for the CZ and the UWBZ. The AF has not performed the EBR pilot test in the UWBZ that was agreed to in the ST012 Work Plan.
Reported on AF flowchart as Eh
AF decision flowchart only mentions "Iron" as an analyte, without differentiating which iron species will be monitored
AF decision flowchart only mentions "Iron" as an analyte, without differentiating which iron species will be monitored
All items other than the last metric, and using qPCR to determine the size of the sulfate-reducing population, are included as part of the already-proposed standard stable-isotope probe (SIP; Bio-Trap) study listed on the AF decision flowchart, but are not included in the metrics to be reported. All of these data are key to fully understanding the makeup, activities, and health of the indigenous microbial population.
These samplers cannot be used in LNAPL, but can be deployed underneath LNAPL.

Total size of sulfate-reducing bacteria population	Υ
Total size of benzene-degrading	
bacteria population	
In-situ benzene degradation rate	
Amount of benzene converted to	Υ
biomass during stable isotope study	
Amount of benzene converted to	
carbon dioxide during stable isotope	Υ
study	
The overall health of the indigenous	
microbial population, as determined via	
PLFA analyses	
The dominant electron-accepting	
process for indigenous microbial	
population, and reason for the	
conclusion	

#### Assessments During EBR

# **Hydrogeologic Data**

Groundwater gauge data (depth to	
water, depth to product, product	
thickness)	
Biofouling	Υ

# **Mapping Contaminant Locations and Concentrations**

Locate and map LNAPL presence and depth - monitoring wells	У
Locate and map dissolved-phase benzene presence and concentration	у
Locate and map dissolved-phase VOC presence and concentration	У
Calculate total LNAPL mass	
Determine the content of COCs in the LNAPL	

		New and existing MWs, located in the area to be impacted by injections/ amendments, and downgradient of this area
	quarterly	
During EBR		All new and existing MWs, located in the area to be impacted by injections/amendments, and downgradient of this area
	Timing of sampling and analysis to follow schedule outlined in Table 4.1 of referenced document; mapping performed once per month	
	Quarterly Quarterly	MWs with recoverable NAPL located in the area to be impacted by injections/ amendments

These assessments will be used to monitor the progress of
EBR, and to determine if changes to the EBR strategy need to
be made. These will also help monitor progress of EBR.
Comparison of NAPL compositions before/during EBR to assess
Comparison of NAPL compositions before/during EBR to assess reductions in COC content

qPCR performed in addition to the stable-isotope study. AF decision flowchart references SRB gene, but Microbial
Insights uses the APS gene to screen for sulfate reducers. Unclear as to what "SRB" gene is being referenced in
flowchart.
Synoptic measurements should be made to allow accurate development of hydraulic head maps and evaluation of
groundwater to produce gw flow directions
See AF Decision Tree
See Al Decision free
Need to ensure good knowledge of locations where EBR treatments/amendments are being conducted, as well as
downgradient. Timing schedule found in: Final Field Variance Memorandum #5 – Extraction and Treatment System
Construction, Former Liquid Fuels Storage Area, Site ST012, Former Williams Air Force Base, Mesa, Arizona; 01 Dec 2016
Measurements of NAPL content, specifically benzene mole fraction, are a primary parameter for assessing EBR
performance. See the "Figures" tab for example plots of benzene mole fraction. Refer to other comments in
"modeling" sections of this table.

and map sulfate concentrations argeted treatment area as well Y ngradient
a time estimate for sufficient epletion in LNAPL, groundwater,
details of EBR modeling to
e time estimates for
ation
proof of concept supporting the
reduction for EBR
details used to determine the sulfate injection strategy.
Surface injection strategy.
ature
Υ
ue Y
ed Oxygen Y
Y
orus
Iron
on
Y

During EBR	At least annually	
	Monthly for the first	
During EBR	quarter of EBR, followed	New and existing MWs
	by quarterly	

Demonstrate achievement of remediation goals based on observed benzene concentration reductions in <u>LNAPL and groundwater</u> .  Modeling and analyses of field data should also incorporate
geochemical (e.g., sulfate) and microbial data (e.g., biomass) parameters that support hydrocarbon mineralization by
biodegradation mechanisms (separate from dilution or sorption
mechanisms). Modeling needs to evaluate rate-limited dissolution
of LNAPL constituents so that the extent to which benzene and
other hydrocarbon concentration reductions in groundwater are
due to slow NAPL/aqueous-phase mass transfer (refer to example
calculations in "Figures" tab). Sensitivity analyses should also be
performed to rigorously document the variability of remediation
timeframes as a function of EBR parameters.
Ta hada magaitan harrantanahista manistrat anati 190
To help monitor key microbial nutrient availability
Will help determine preferred TEA for indigenous microbes
The state of the s
Will help determine preferred TEA for indigenous microbes
To monitor if periodic sulfate injections or recirculation are
necessary to sustain degradation rates
necessary to sustain degradation rates

When compared to this baseline data, this information will help monitor sulfate migration outside of the COC areas
Please provide a detailed evaluation of important factors determining the efficiency and rate of COC biodegradation and depletion of COCs from the LNAPL source materials. In this evaluation, sensitivity analyses should be incorporated for evaluating the effect of assumptions on remedial efficacy and timeframe scenarios. This will aid in evaluating remedy effectiveness. Please use EPA-team modeled Time of Remediation estimates provided on May 30, 2017 as an example of the detail that should be provided. ST12 Joint agency EBR model cover letter.pdf; TOR Estimates_ST012_052217.pdf; BIONAPL_Box_Model_revised_04-27-2017_UWBZ.xls].
Please use EPA-generated models as an example of sufficient documentation, to allow for an independent evaluation of the results of the modeling. Please see comments provided in: Techlaw memo dated March 24, emailed on April 11, 2017, and Time of Remediation estimates provided on May 30, 2017 (ST12 Joint agency EBR model cover letter.pdf; TOR Estimates_ST012_052217.pdf; BIONAPL_Box_Model_revised_04-27-2017_UWBZ.xls])
Ongoing updates as field data become available
These analyses will provide an indirect method of monitoring the indigenous microbial community.
Reported on AF flowchart as Eh
AF decision flowchart only mentions "Iron" as an analyte, without differentiating which iron species will be monitored  AF decision flowchart only mentions "Iron" as an analyte, without differentiating which iron species will be monitored

	Hydrogen Sulfide	
	Methane	
	Alkalinity	
	TPH (DRO, GRO)	Υ
	VOCs	Υ
	Arsenic	Υ
TEA Injection F	luid	
	ICP Metals	Υ
	Details of injection material composition	
	Sulfate	Υ
	Location of each injection/amendment	
	Concentration of sulfate at each	
	injection/ amendment location	
	Anticipated zone of influence for each	
	injection/ amendment	
Indigenous Mid	crobial Population	
	Total size	
	Major groups within population, and	
	their proportion of total	
	Total size of sulfate-reducing bacteria population	Υ
	Total size of sulfate-reducing bacteria	Y
	Total size of sulfate-reducing bacteria population	Y

Amount of benzene converted to

biomass during stable isotope study

Υ

During EBR, for every injection/ amendment event and location		
	Monthly, per Table 5.1 Need to check each batch	
During EBR, 6-9 months post-injection (per Decision Matrix)	At least once during EBR, 4-6 weeks after initial sulfate injection. May need to be repeated if geochem data suggests a problem.	Samplers should be placed so as to monitor the core of sulfate injections, its periphery, and downgradient.  All three zones should be monitored.  The same wells should be monitored pre-EBR, during EBR, and post-EBR.

To monitor if hydrogen sulfide concentrations inhibit degradation or will subsurface conditions mitigate their buildup?
To record makeup and concentration of injection fluid
Will the injected sulfate become well distributed with respect to NAPL accumulations?
These analyses will quantify the size, makeup, and health of the indigenous microbial community.
If there are indications that the microbial population is struggling during EBR, the analyses should be repeated to determine if alternate strategies are needed
May also help determine lag time for SRBs to acclimate to elevated sulfate concentrations and determine if highly concentrated injections of sulfate will be inhibitive to bacterial activity

T T
Taken from Table 5.1, RD-RAWP Addendum 2 (March 2016); This data will provide a record of exactly what was
injected, where, and at what concentration. This, when compared with the response by the contaminants and other
geochemical and biological data, will help determine if any changes need to be made to amendment variables such as
frequency, concentration, etc.
Any metals over MCL would prevent ability to inject
This may be proprietary; however, an effort to obtain this information should be made
Need to check the TEA fluid before injection fluid before goes into ground to ensure that the concentration is as expected , was mixed and diluted correctly, etc.
All items other than the last metric, and using qPCR to determine the size of the sulfate-reducing population, are
included as part of the already-proposed standard stable-isotope probe (SIP; Bio-Trap) study listed on the AF decision
flowchart, but are not included in the metrics to be reported. All of these data are key to fully understanding the
makeup, activities, and health of the indigenous microbial population.
makeup, activities, and nearth of the margenous interoblar population.
There are also are the constitution of the same has dead and an also also the INIADI
These samplers cannot be used in LNAPL, but can be deployed underneath LNAPL.
Taken from Table 5.1, RD-RAWP Addendum 2 (March 2016). AF decision flowchart references SRB gene, but Microbial
Insights uses the APS gene to screen for sulfate reducers. Unclear as to what "SRB" gene is being referenced in
flowchart. qPCR performed in addition to the stable-isotope study.

	Amount of benzene converted to carbon dioxide during stable isotope study	Υ
	The overall health of the indigenous microbial population, as determined via PLFA analyses	
	The dominant electron-accepting process for indigenous microbial population, and reason for the conclusion	
ost-EBR Data		
Hydrogeolog	gic Data	
	Groundwater gauge data (depth to water, depth to product, product thickness)  Biofouling	Υ
Manning Co	entaminant Locations and Concentration	
Mapping Co	The content and concentration	
	Locate and map LNAPL presence and	
	depth	
	Locate and map dissolved-phase benzene presence and concentration, in	
	excess of 5 ug/L	
	Locate and map dissolved-phase VOC	
	presence and concentration	
	Calculate total LNAPL mass present	
	Determine the content of COCs in the	

		Each MW used for injections, amendments,
Post-EBR		or any analyses
	Minimum of semi- annual	
	once	
Post-EBR	Quarterly, then frequency amended per	
	modeling and EPA	or any analyses
	modeling and EPA guidance on MNA	or any analyses
		MWs with recoverable NAPL located in the area to be impacted by injections/

This data will be compared against baseline data, and data taken during EBR, to determine the success of the project as well as to	
dentify necessary future actions. This data will also become the baseline information used at the start of MNA	
o ensure no biofouling after EBR	
o choure no protoculing after EDI	
Comparison of NAPL compositions before/during/after EBR to assess reductions in COC content	

EBR remedial goals include:
1) Depletion of COC concentrations (mole fractions) in LNAPL to the degree that the COC-depleted LNAPL cannot transfer COCs to groundwater above MCLs
2) Reduction of COC concentrations in site groundwater and soil to the degree that MNA could be expected (based on Regulatory Agency-approved modeling) to reduce COCs in groundwater below MCLs within the ROD remedial timeframe.
Specific numerical metrics, milestones, and timelines (i.e., specific concentrations of COCs in LNAPL and groundwater, along with associated geochemical and microbiological data, at specific times after initial implementation of EBR, and of MNA) will be developed based on Regulatory Agency - approved modeling efforts to guide remedial activities, evaluate success of the remedial approaches, and trigger contingency remedies if necessary.
Synoptic measurements should be made to allow accurate development of hydraulic head maps and evaluation of groundwater to produce gw flow directions
Pope, Daniel F., Steven D. Acree, Herbert Levine, Stephen Mangion, Jeffrey van Ee, Kelly Hurt, Barbara Wilson, Performance Monitoring of MNA Remedies for VOCs in Ground Water EPA/600/R-04/027, National Risk Management Research Laboratory Office Of Research And Development U.S. Environmental Protection Agency, Ada OK, 2004
Update based on additional field data
Measurements of NAPL content, specifically benzene mole fraction, are a primary parameter for assessing EBR performance. See the "Figures" tab for example plots of benzene mole fraction. Refer to other comments in "modeling" sections of this table.

	Locate and map sulfate concentrations in the targeted treatment area as well as downgradient	Υ
Modeling		
	Provide a time estimate for sufficient COCs depletion in LNAPL, groundwater, and soil by MNA	
	Provide details of post-EBR modeling to	
	calculate time estimates for remediation	
GW Geocher	remediation	
GW Geocher	remediation	Y Y

Temperature	Y
pH	Υ
ORP value	Υ
Dissolved Oxygen	Υ
Nitrate	Υ
Phosphorus	
Ferrous Iron	
Total Iron	
Sulfate	Υ
Hydrogen Sulfide	
Methane	
Alkalinity	
TPH (DRO, GRO)	Υ
VOCs	Υ
Arsenic	Υ

Post-EBR	As needed	
Post-EBR	Quarterly, then frequency amended per modeling and EPA guidance on MNA	Each MW used for injections, amendments, or any analyses

Demonstrate achievement of remediation goals based on observed benzene concentration reductions in <a href="LNAPL">LNAPL</a> and groundwater. Modeling and analyses of field data should also incorporate geochemical (e.g., sulfate) and microbial data (e.g., biomass) parameters that support hydrocarbon mineralization by biodegradation mechanisms (separate from dilution or sorption mechanisms). Modeling needs to evaluate rate-limited dissolution of LNAPL constituents so that the extent to which benzene and other hydrocarbon concentration reductions in groundwater are due to slow NAPL/aqueous-phase mass transfer (refer to example calculations in "Figures" tab). Sensitivity analyses should also be performed to rigorously document the variability of remediation timeframes as a function of EBR parameters.

When compared to this baseline data, this information will help monitor sulfate migration outside of the COC areas
Please provide a detailed evaluation of important factors determining the efficiency and rate of COC biodegradation and depletion of COCs from the LNAPL source materials. In this evaluation, sensitivity analyses should be incorporated for evaluating the effect of assumptions on remedial efficacy and timeframe scenarios. This will aid in evaluating remedy effectiveness. Please use EPA-team modeled Time of Remediation estimates provided on May 30, 2017 as an example of the detail that should be provided. ST12 Joint agency EBR model cover letter.pdf; TOR Estimates_ST012_052217.pdf; BIONAPL_Box_Model_revised_04-27-2017_UWBZ.xls].
Pope, Daniel F., Steven D. Acree, Herbert Levine, Stephen Mangion, Jeffrey van Ee, Kelly Hurt, Barbara Wilson, Performance Monitoring of MNA Remedies for VOCs in Ground Water EPA/600/R-04/027, National Risk Management Research Laboratory Office Of Research And Development U.S. Environmental Protection Agency, Ada OK, 2004
Reported on AF flowchart as Eh
13p 2 3 2 3 3 1 1 1 1 2 1 2 1 2 1 2 1 2 1 2
AF decision flowchart only mentions "Iron" as an analyte, without differentiating which iron species will be monitored
AF decision flowchart only mentions "Iron" as an analyte, without differentiating which iron species will be monitored

# **Indigenous Microbial Population**

Total size	
Major groups within population, and	
their proportion of total	
Total size of sulfate-reducing bacteria	Υ
population	
Total size of benzene-degrading	Υ
bacteria population	, ř
In-situ benzene degradation rate	
Amount of benzene converted to	Υ
biomass during stable isotope study	
Amount of benzene converted to	
carbon dioxide during stable isotope	Υ
study	
The overall health of the indigenous	
microbial population, as determined via	
PLFA analyses	
The dominant electron-accepting	
process for indigenous microbial	
population, and reason for the	
conclusion	

Post-EBR	Once, within 3 months of the last injection/ amendment	Samplers should be placed so as to monitor the core of sulfate injections, its periphery, and downgradient.  All three zones should be monitored.  The same wells should be monitored pre-EBR, during EBR, and post-EBR.

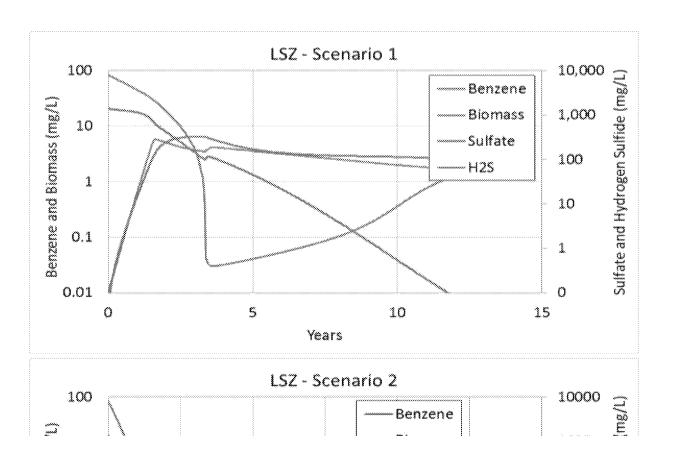
These analyses will quantify the size, makeup, and health of the indigenous microbial community at the end of EBR, and will provide baseline data for MNA

All items other than the last metric, and using qPCR to determine the size of the sulfate-reducing population, are included as part of the already-proposed standard stable-isotope probe (SIP; Bio-Trap) study listed on the AF decision flowchart, but are not included in the metrics to be reported. All of these data are key to fully understanding the makeup, activities, and health of the indigenous microbial population.
These samplers cannot be used in LNAPL, but can be deployed underneath LNAPL. The use of the stable-isotope probes would be anticipated as a one-time event, unless groundwater data suggests a need to perform it again.
AF decision flowchart references SRB gene, but Microbial Insights uses the APS gene to screen for sulfate reducers. Unclear as to what "SRB" gene is being referenced in flowchart. qPCR performed in addition to the stable-isotope study.

Example calculations based on scenarios described in "Time of Remediation Estimates, Enhanced Bioremediation at ST01 Calculation input is provided in Tables 8-10 of the TOR memorandum

Table 8. Parameters for Monod Kinetics

Parameter	_	UWBZ	LSZ	Reference
Vsoii	γd^3	122,556	38,500	Table 2
Q	gpm	4.4	3.5	Table 2
Knapi	1/day	0.05	0.05	Mobile et al. (2016)
C <sup>02</sup> (backgrnd)	mg/L	7.0	7.0	Table M.4.3.2.1
C <sup>NO3-</sup> (backgrnd)	mg/L	8.0	8.0	Table M.4.3.2.1
C <sup>504-</sup> (backgrnd)	mg/L	200	290	Table M.4.3.2.1
<sub>7/</sub> 504-	g/g	4	4	Table M.4.3.5.3
v max Benzene,502-	1/day	0.000875	0.0175	Table M.4.3.5.1/2
V <sup>max</sup> Toluene,SO <sub>4</sub> <sup>2-</sup>	1/day	0.001125	0.0225	Table M.4.3.5.1/2
ymax Ethylbenzene,502	1/day	0.000875	0.0175	Table M.4.3.5.1/2
ymax vvienes sof	1/day	0.001125	0.0225	Table M.4.3.5.1/2
max VNaphthalene,502 <sup>2</sup>	1/day	0.000125	0.0025	Table M.4.3.5.1/2
ymax TMB.SO <sub>4</sub> 2	1/day	0.000125	0.00125	Table M.4.3.5.1/2
ymax Other Aromatics,50‡	1/day	0.000625	0.0125	Table M.4.3.5.1/2
$K_{SO_4^{2-}}$	mg/L	1	1	Table M.4.3.5.3
K, 503.	mg/L	5	5	Table M.4.3.5.3
Y	g/g	0.2	0.2	BEM (2007)
Msea,o (initial)	mg/L	0.01	0.01	BEM (2007)
ld.bk lsrb	1/day	0.001/0.0	0.001/0.0	BEM (2007)



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		`_
Aquifer Zone		4
		1
		F
UWBZ	NAPL (gal)	
V = 122,556 cy	Sulfate (kg) =	
	Sulfate (mg/L) =	
LSZ	NAPL (gal)	
V = 38,500 cy	Sulfate (kg) =	
	Sulfate (mg/L) =	

Table 10. TOR for NAPL Deple

ianie to: iovioi imarr nehie					
Aquifer	Ambient	Mass	Calci		
Zone	Flow	Transfer	Targe		
		Coeff.	Vol		
			Poros		
	gpm	day¹	yε		
UWBZ	4.4	0.0042	1		
UW8Z	4.4	0.05	ķ		
UW8Z	0.0*	0.05	1		
LSZ	3.5	0.0042	5		
LSZ	3.5	0.05	1		
LSZ	0.0*	0.05	1:		

Targeted Sulfate Mass and Concentration

rai Peren annare mass and concernation						
Calculated^	Calculated^	Literature*	Literature*			
Γarget NAPL	Target NAPL	Target NAPL	Target NAPL			
Volume	Volume	Volume	Volume			
Porosity=0.3	sity=0.3 Porosity=0.4 Porosity=0.3		Porosity=0.4			
gal	gal	gal	gal			
250,999	215,142	294,399	395,887			
1,032,067	884,629	1,210,521	1,627,823			
36,715	23,603	43,064	43,432			
54,821	46,989	110,682	155,783			
225,415	193,211	455,106	640,554			
25,527	16,410	51,538	54,404			



Trees assers message as constant and a second second second						
ulated	Calculated	Literature	Literature	Notes		
t Napl	Target NAPL	Target NAPL	Target NAPL			
lume	Volume	Volume	Volume			
:ity=0.3	Porosity=0.4	Porosity=0.3	Porosity=0.4			
ears	years	years	years			
:33	111	152	178	1		
92	84	102	126	Ť.		
26	116	140	174	2		
2.4 🕖	36.2	104	116	3		
3.2	9.4	28.0	36.1	m		
2.1	9.9	22.0	27.0	4		

